

Ms Excel version (.xl)

Listed are all species with validly published names except for *A. boissieri* and *A. neofarctii*, which are negative in the assimilation tests shown (they utilize other substrates such as sucrose or D-fructose). Not included are *A. grimontii*, *A. guangdongensis*, *A. pakistanicus*, and *A. dikhonisii* which are the later synonyms of *A. junii* (Vanechouette et al., 2008), *A. indicus* (Nemeć & Radloff-Krizova, 2016), *A. bohemicus* (Nemeć & Radloff-Krizova, 2016), and *A. lactucae* (Dunlap & Rooney, 2018), respectively. Genomic species 6, 15B1 and 16, and Taxon 21 and 22 are provisional designations of putative novel species. Each species includes a type strain. Numbers of strains of individual species are indicated in parentheses after the species names. The results [except for those for *A. gracilis* and *A. portensis* adopted from Carvalheira et al. (2020), *A. chengduensis* from Qin et al. (2020), and *A. lanii*, *A. shaoyingense*, and *A. wangaense* from Zhu et al. (2020)] were obtained in the Laboratory of Bacterial Genetics in Prague using an array of standardized, in-house tests as described by Nemeć et al. (2009) and Krizova et al. (2015), and were published previously (Hu et al., 2018; Krizova et al., 2015; Nemeć et al., 2017, 2018; Qin et al., 2019) except for the unpublished results for Taxons 21 and 22. Except for temperature-dependent tests, the culture temperature was 25 °C for *A. celiucus* and 30 °C for the other species. Assimilation/growth tests were interpreted after 6 days of culture, other tests after 3 (amylase and gelatinase activities) or 2 (D-glucose acidification, temperature growth tests) days. +, All strains positive; -, all strains negative; ?, results are opposite to those published in original nomenclatural proposals; D, (mostly) doubtful or irreproducible reactions; W, (mostly) weak positive reactions; ND, not determined. Numbers are percentages of strains with clearly positive reactions; for strain-dependent reactions, results for type strains are given in parentheses.

References

- Carvalho A, González-Siles L, Salva-Serra F et al. (2020) *Acinetobacter portensis* sp. nov. and *Acinetobacter guevara* sp. nov., isolated from raw meat. *Int J Syst Evol Microbiol* 70: 4544–4554.

Dunlap CA & Rooney AP (2018) *Acinetobacter dijkshoorniae* is a later heterotypic synonym of *Acinetobacter luteus*. *Int J Syst Evol Microbiol* 68: 131–132.

Hu Y, Feng Y, Qin J et al. (2018) *Acinetobacter wuhaeensis* sp. nov., isolated from hospital sewage. *Int J Syst Evol Microbiol* 68: 3212–3216.

Krizova L, Makriliaeva M, Sedo M & Nemec A (2015) *Acinetobacter albensis* sp. nov., isolated from natural soil and water ecosystems. *Int J Syst Evol Microbiol* 65: 3905–3912.

Nemec A & Radulovaca-Krizova L (2016) *Acinetobacter pakistanensis* Abbas et al. 2014 is a later heterotypic synonym of *Acinetobacter bohemicus* Krizova et al. 2014. *Int J Syst Evol Microbiol* 66: 5614–5617.

Nemec A & Radulovaca-Krizova L (2017) *Acinetobacter quangdongensis* Feng et al. 2014 is a further heterotypic synonym of *Acinetobacter indicus* Malhotra et al. 2012. *Int J Syst Evol Microbiol* 67: 4080–4082.

Nemec A, Makriliaeva M, Radulovaca L et al. (2019) *Acinetobacter bejenensis* sp. nov. and *Acinetobacter gyllenbergii* sp. nov., haemolytic organisms isolated from humans. *Int J Syst Evol Microbiol* 59: 118–124.

Nemec A, Makriliaeva M, Radulovaca L et al. (2020) Revising the taxonomy of the *Acinetobacter lwoffii* group: the description of *Acinetobacter pseudowolffii* sp. nov. and emended description of *Acinetobacter lwoffii*. *Syst Appl Microbiol* 42: 159–167.

Nemec A, Radulovaca-Krizova L, Makriliaeva M & Sedo O (2017) *Acinetobacter colistimotolerans* sp. nov. (formerly genomic species 13 sensu Bouvet & Jeanjean and genomic species 14 sensu Tjøberg & Ussing), isolated from human infections and characterised by intrinsic resistance to polymyxins. *Int J Syst Evol Microbiol* 67: 2134–2141.

Qin J, Feng Y, Lu X & Zong Z (2020) *Acinetobacter chengduensis* sp. nov., isolated from hospital sewage and capable of acquisition of carbapenem resistance genes. *Syst Appl Microbiol* 43: 126092.

Qin J, Hu Y, Feng Y et al. (2018) *Acinetobacter schinasiensis* sp. nov., recovered from hospital sewage in China. *Int J Syst Evol Microbiol* 68: 3897–3901.

Qin J, Makriliaeva M, Nemec M et al. (2019). Description of *Acinetobacter curvulans* sp. nov., isolated from hospital sewage and capable of acquisition of multiple antibiotic resistance genes. *Syst Appl Microbiol* 2019: 42: 319–325.

Vaneeschoutte M, De Baere T, Nemec A et al. (2008) Reclassification of *Acinetobacter grimontii* Carr et al. 1986 as a later synonym of *Acinetobacter junii* Bouvet and Grimont 1986. *Int J Syst Evol Microbiol* 58: 937–940.

Zhu W, Dong K, Yang J et al. (2020) *Acinetobacter lanai* sp. nov., *Acinetobacter shadymingi* sp. nov. and *Acinetobacter wanghaiense* sp. nov., isolated from faeces of *Equus kiang*. *Int J Syst Evol Microbiol* doi:10.1099/ijsem.0.004567 (in press)